

GeneFinder™ COVID-19 PLUS

RealAmp Kit

Instructions for Use

REF

IFMR-45

IVD



REF

IFMR-45

**Manufacturer:****OSANG Healthcare Co., Ltd**

132, Anyangcheondong-ro, Dongan-gu, Anyang-si, Gyeonggi-do, 14040 Korea

Tel: +82-31-460-9937

Fax: +82-31-460-9933

Web site: <http://www.osanghc.com>

EC REP

Obelis S.A

Bd. General Wahis 53,1030 Brussels, Belgium

Tel: +32-2-732-59-54

Fax: +32-2-732-60-03

E-mail: mail@obelis.net**In Vitro Diagnostic**

1. Intended Use

GeneFinder™ COVID-19 PLUS RealAmp Kit is used for detection of COVID-19 (COIVD-19) virus through reverse Transcription and Real-Time Polymerase Chain Reaction from RNA extracted from Respiratory specimens such as Alveolar lavage fluid, throat swab, sputum. This product can qualitatively detect COVID-19 using Polymerase Chain Reaction.

2. Principle of the Assay

One-Step Reverse Transcription Real-Time polymerase chain reaction is used to confirm the presence of COVID19 by amplification of RdRp, E and N gene

This product is a In vitro diagnostics (IVD) and is used by professionals in hospitals and laboratories.

3. Kit Contents

| Reagents / Materials | 100 tests/Kit |
|--------------------------------|---------------|
| COVID-19 PLUS Reaction Mixture | 1,050 ul |
| COVID-19 PLUS Probe Mixture | 550 ul |
| COVID-19 PLUS Positive Control | 50 ul |
| COVID-19 PLUS Negative Control | 50 ul |

4. Storage and Handling Requirements

- All components of the kit should be stored at -20°C or below and kept stable until the expiry date stated on the label.
- The COVID-19 PLUS Probe Mixture must be stored below -20°C and in the dark.
- Expires 12 months after date of manufacture. Do not use after expiration date.
- Expires 6 months after opening the kit. Do not use after use life time.
- Store the rest of the kit below -20°C.
- If the kit is defective, do not use.
- Dispose of unused reagents and waste in accordance with country, federal, state and local regulations.

Note: Inaccurate results can be obtained if the kit is stored at room temperature for a long period of time.

Note: Unnecessary repeated freezing and thawing lead to inaccurate results.

5. Product Description

1) COVID-19 PLUS Reaction Mixture

COVID-19 PLUS reaction mixture with reagents for reverse transcription and amplification

2) COVID-19 PLUS Probe Mixture

Buffer solution with specific primers and probes which conjugate with nucleic acids of COVID-19 virus and internal control

3) COVID-19 PLUS Positive Control

Positive Control determines for presence of errors/contamination during the test.

Caution. Care should also be taken to avoid cross-contamination of other samples when adding Positive Control.

4) COVID-19 PLUS Negative Control

To confirm the absence of contamination, negative control reaction should be included at every run as it indicates that reagents have not been contaminated.

* The product allows the accomplishment of 100 tests, including controls.

6. Required materials

6.1. Provided in the product

(100 tests / Kit)

| Label | Cap color | Storage | Quantity |
|--------------------------------|-----------|---------|--------------|
| COVID-19 PLUS Reaction Mixture | Purple | -20°C | 1 x 1,050 ul |
| COVID-19 PLUS Probe Mixture | Brown | -20°C | 1 x 550 ul |
| COVID-19 PLUS Positive Control | Red | -20°C | 1 x 50 ul |
| COVID-19 PLUS Negative Control | Green | -20°C | 1 x 50 ul |

6.2. Required but not provided in the product

- Applied Biosystems® 7500 / 7500 Fast Real Time PCR Instrument System and CFX96 real time PCR system.
- Pipettes (1- 20 µl, 20-200 µl, 200-1,000 µl)
- Pipettes tips with aerosol barrier (RNase, DNase-free)
- Powder-free gloves (disposable)
- Vortex mixer or equivalent
- 1.5 ml tube
- PCR tube or 96 well plate
- Bench microcentrifuge
- RNA isolation kit (Use of QIAamp Viral RNA Mini Kit (Cat. # 52904, Qiagen) is recommended or commercial kits)

7. Warning and Precaution

The GeneFinder™ COVID-19 Plus RealAmp Kit is designed for **In vitro diagnostics**

General warnings and precautions

- Read the instructions in the package carefully before processing samples.
- Use 0.5% v / v sodium hypochlorite or another disinfectant to clean and disinfect the area around the sample.
- Decontaminate and dispose of all specimens, reagents and other potentially contaminated materials in accordance with local regulations.
- Use universal precautions when performing the assay. Handle samples as if capable of transmitting infection.
- Wear personal protective apparel, including disposable gloves, throughout the assay procedure. Thoroughly wash hands after removing gloves, and dispose of gloves as biohazardous wastes.
- The material that come into contact with the biological samples must be autoclaved for one hour at 120°C before disposal.
- Do not eat, drink, smoke, or apply cosmetics in areas where reagents of samples are handled.
- Do not pipet by mouth.
- Do not use a kit after its expiration date.

- Use aerosol-resistant pipette tips and use a new tip every time a volume is dispensed.
- Store the reagents recommended temperature.
- Do not mix reagent from different batches of the kit.
- Store the kit away from any source of contaminating DNA, especially amplified nucleic acid. - Use sterile disposable laboratory materials and do not re-use the tubes and tips.
- Alterations in the physical appearance of kit components may indicate instability or deterioration.
- Use all pipetting devices and instruments with care and follow the manufacturer's instructions for calibration and quality control.
- Do not modify the reagent/sample volume used in the test or use in a wrong way which is not recommended.
- Store COVID-19 PLUS Probe Mixture at -20°C in a dark place.

8. Procedure

8.1 Preparation of sample

The GeneFinder™ COVID-19 PLUS RealAmp Kit must be used with RNA extracted from Alveolar lavage fluid, throat swab, and sputum samples. RNA extraction is recommended from sample as soon as possible for accurate experiments.

8.1 RNA Extraction

Commercialized extraction kit should be used for collection of RNA Extracted samples. QIAamp viral RNA Mini Kit (Qiagen, Germany, Cat. # 52904) is recommended for extraction. Please carry out RNA extraction according to the manufacturer's instructions.

Extracted RNA samples are more vulnerable than DNA that it is suggested to avoid repeated freezing and thawing and to keep at -70°C.

When extracting RNA, be sure to extract it according to the manufacturer's instructions.

8.2 Preparation of reagents

Thaw all components thoroughly at room temperature before using. Mix gently, spin down the contents for 5 seconds, and then test it immediately..

1. Mix 10 µl of COVID-19 PLUS Reaction Mixture, 5 µl of COVID-19 PLUS Probe Mixture to prepare RT-PCR Master mixture as described in the following table (Table 1). Prepare enough volume of Master mixture for all the reactions plus extra amounts to prevent possible pipetting error.

Note: Total Master mixture number

= n sample + 1 positive control + 1 negative control + 1 extra

| Solution | Number of Samples | | | Total volume of Master Mixture |
|--------------------------------------|-------------------|--------|--------|--------------------------------|
| | 1 TEST | 3 TEST | 5 TEST | |
| COVID-19 PLUS Reaction Mixture | 10 ul | 30 ul | 50 ul | 10x(n+3) |
| COVID-19 PLUS Probe Mixture | 5 ul | 15 ul | 25 ul | 5x(n+3) |
| Total (COVID-19 PLUS Master Mixture) | 15 ul | 45 ul | 75 ul | 20x(n+3) |

Table 1. Master Mixture preparation

Important: Adequate controls should be used in each run to ensure reliable results.

2. Place 15 µl of RT-PCR Master mixture into each PCR tube or optical 96 well plate **Note:** To avoid any bubbles, do not vortex the tubes at this step.
3. Add each of 5 µl of sample RNA into the corresponding PCR tube/well-plate for amplification and mix them with pipetting

4. Place 5 µl of Positive Control and Negative Control into the each PCR tube or well-plate in the same way.
Note: Total Reaction volume is 20 µl per sample.
Note: Insufficient mixing of the Master mixture may lead to inaccurate result..
5. Transfer PCR tube or well-plate for testing into the real-time thermal cycler and start the thermal cycle for the amplification.

Figure 1. Schematic workflow for test

8.3 Setting of the RealTime Amplificatino



Prior to amplification, operate PCR instrument according to the manufacturer’s manual.

✂ Real-Time PCR condition

| | Step | Temperature | Time | Cycles |
|---|-----------------------|-------------|--------|-----------|
| 1 | Reverse Transcription | 50°C | 20 min | 1 cycle |
| 2 | Pre-denaturation | 95°C | 5 min | 1 cycle |
| 3 | Denaturation | 95°C | 15 sec | 45 cycles |
| | Annealing* | 58°C | 60 sec | |

*Collection of data

✘ Fluorescence setting

| Target | Fluorescence |
|------------------|-------------------------|
| RdRp gene | FAM |
| E gene | Texas Red |
| N gene | JOE (ABI) / VIC (CFX96) |
| Internal Control | Cy5 |

8.4 Data analysis

1. Select Amplification Plot at Analysis Mode.
2. Select Analysis Settings.
3. Set Threshold Values, Baseline start and end values.

| Target | Threshold | | Baseline | |
|------------------------|-------------------------|--------|----------|-----|
| | ABI 7500/ ABI 7500 fast | CFX 96 | Begin | End |
| RdRp gene (FAM) | 30,000 | 300 | 3 | 15 |
| E gene (Texas Red) | 30,000 | 300 | 3 | 15 |
| N gene (JOE) | 30,000 | 300 | 3 | 15 |
| Internal Control (Cy5) | 10,000 | 100 | 3 | 15 |

9. Result

Result Interpretation.

| # | Ct range | | | | Result |
|---|------------|---------------|---------|----------|--|
| | RdRp (FAM) | E (Texas Red) | N (JOE) | IC (Cy5) | |
| 1 | ≤43 | ≤43 | ≤43 | ≤35* | COVID-19 Positive |
| 2 | ≤43 | ≤43 | U.D | ≤35 | |
| 3 | ≤43 | U.D | ≤43 | ≤35 | |
| 4 | ≤43 | U.D | U.D | ≤35 | Repeat the test (COVID-19 Positive if RdRp≤43) |
| 5 | U.D | ≤43 | ≤43 | ≤35 | Repeat the test (COVID-19 Positive if E and N≤43) |
| 6 | U.D | U.D | ≤43 | ≤35 | Repeat the test (COVID-19 Positive if N≤43) |
| 7 | U.D | ≤43 | U.D | ≤35 | Beta coronavirus |
| 8 | U.D | U.D | U.D | ≤35 | Negative |
| 9 | U.D | U.D | U.D | U.D | Invalid (re-test) |

Note

- *When target RNA is detected in a sample amplification reaction, Internal control (IC) may give the result as Ct Not applicable (N/A). In fact, low-efficiency amplification reaction for internal control may be displaced by competition from high-efficiency amplification reaction for Target gene. In such a case, it shall be determined as positive.
- If test result is not valid as follows, it is recommended to retest.
 - ① In case of Ct value of Internal control is Not applicable
 - ② In case of Invalid result.

10. Quality Control

Validating the whole analysis procedure (of each extraction and amplification session by processing a negative tested sample and a positive tested sample or a calibrated reference material) is recommended.

11. Procedure Limitation

- The users must be trained and familiar with this technology prior to the use of this device.
- Any diagnostic results generated must be interpreted in conjunction with other clinical or laboratory findings. It is the user's responsibility to validate system performance for any procedures used in their laboratory which are not covered by the OSANG Healthcare performance studies.
- Use this product only with RNA extracted from the following human biological samples: Alveolar lavage fluid, throat swab, sputum
- A negative result does not exclude the possibility of infection, because results are dependent on appropriate specimen collection and absence of inhibitors. The presence of PCR inhibitors may cause invalid results with this product..

12. Trouble shooting

| Problems | Possible Causes | Recommendation |
|---|--|--|
| If no fluorescent signal is detected in all samples, including positive control | Error in the preparation of the master mixture | Check the volumes of reagent dispensed during preparation of the master mixture |
| | Inhibitors added | Take care when RNA is extracted and repeat the extraction step with new sample |
| | Probe degradation | Use a new probe aliquot |
| | Positive control degradation | Use a new aliquot of Positive control |
| | Omitted components | Verify each component and repeat the PCR mixture preparation |
| | Instrument setting error | Check position settings for the positive control on the instrument Check the thermal cycle settings on the instrument |
| If the fluorescent signal is detected in negative control reaction except CY 5. | Carry-over contamination | Take care when dispensing samples, negative controls, and positive controls on the instrument Always change tips between one sample and another |
| | Microplate/ tube Error | Be careful not to spill the contents of the tube or plate. |
| | Tube cap badly sealed | Take care when sealing the tube cap |
| | Contamination of the amplification mix | Use a new aliquot of amplification mix |
| | Contamination of the extraction/preparation area for amplification reactions | Clean surfaces and instruments with aqueous detergents, wash lab coats replace test tubes and tips in use |
| If the fluorescent intensity is weak | Poor quality of RNA samples | Extract RNA from samples using the recommended kit, and store the extracted RNA at -70°C |

| | | |
|---|---|---|
| or does not appear only in the unknown samples | Not enough volume of RNA samples added | Repeat the PCR reaction using the correct volume of RNA samples |
| If the fluorescent intensity is weak or does not appear in the positive control | Probe degradation | Use a new probe aliquot Test should be done with a new kit |
| If the diverse intensity of fluorescent signals appear | Pipetting error | Make sure that an equal volume of reactants is added in each tube |
| | Contamination in the outer surface of PCR tubes and plate | Wear gloves during the experiment |

13. Performance characteristics

Analytical Sensitivity : LOD

Analytical sensitivity with Target RNA was performed with GeneFinder™ COVID-19 PLUS RealAmp Kit. As a result of experiments at various concentrations, the analytical sensitivity of the product was confirmed as **10 copies / reaction** for all target genes (RdRp, E, N gene).

Analytical Specificity : Cross Reactivity

The cross reactivity tests were performed using COVID-19 Standard materials and RNA of 14 negative reference materials; None of negative reference materials were detected in any of the test performed.

| # | Name |
|----|-----------------------------------|
| 1 | Influenza A (H1N1/09) |
| 2 | Influenza A (H3N2) |
| 3 | Influenza A (H5N1) |
| 4 | Influenza B |
| 5 | Rhinovirus |
| 6 | Respiratory syncytial virus (A/B) |
| 7 | Parainfluenza 1 virus |
| 8 | Parainfluenza 2 virus |
| 9 | Parainfluenza 3 virus |
| 10 | Parainfluenza 4 virus |
| 11 | Adenovirus |
| 12 | Human Bocavirus |
| 13 | Measles virus |
| 14 | Mycoplasma spp. |

Reproducibility (Between Lots, operators, and places)

Reproducibility between lots, inspectors, and test sites with different manufacturing dates for the same concentration of standard material was found to be within the CV% titration range (<5 CV%) and showed reproducibility.









Repeatability

Repeated every 2 days, the test is carried out by replication; all results are taken as the reference value (5% sustained CV basis).

Interference

Mucin, NaCl, Blood, Respiratory syncytial virus A, and PBS were repeated four times for each of high, medium and low concentrations. As a result of testing with RNA extracted from swab containing interfering substances, it was confirmed that some substances above a certain concentration act as interfering agents in reverse transcription and PCR reactions. At the appropriate concentration, all showed positive test results of 100% and confirmed that they were not affected by the interference materials.

14. Symbols used on Labels

| | | | |
|---|------------------------------------|---|---|
|  | Lot or batch number |  | Caution |
|  | Catalogue number | Store below temperature |  shown |
|  | In Vitro Diagnostic Medical Device |  | Expiry date |
|  | Consult Instruction For Use | Manufacturer |  |

15. Reference

1. WHO COVID-19 report 2020