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Manual

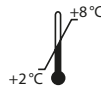
Retinol-binding protein RBP / RBP4 ELISA

*For the in vitro determination des RBP/RBP4
in plasma, serum and urine*

Valid from 2019-03-27



K 6120



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1. INTENDED USE

This Immundiagnostik AG assay is an enzyme immunoassay intended for the quantitative determination of **free retinol-binding protein RBP/RBP4** as well as **RBP4 complexed with transthyretin** in plasma, serum and urine.

For *in vitro* diagnostic use only.

2. INTRODUCTION

Retinol-binding protein RBP/RBP4 is a small (21 kD) transport protein for vitamin A which forms a complex with prealbumin in blood, but loses its affinity for prealbumin once the vitamin has been delivered to the target cells. The free RBP/RBP4 molecule is rapidly filtered at the glomerulus and catabolised in the renal tubules after resorption by the proximal tubular cells (like other small molecules, e.g. β -2 microglobulin). In kidney disease with prevailing tubular changes, these proteins are not reabsorbed and appear in the urine.

As published by Yang et al. (2005), the retinol-binding protein **RBP/RBP4** seems to play a key role in the development of insulin resistance. The fat cell derived peptide RBP/RBP4 also modulates the glucose homeostasis and impairs the insulin sensitivity as well as insulin resistance. The elevation of serum RBP/RBP4 causes systemic insulin resistance, whereas its reduction improves the insulin action.

As a conclusion from the results, the authors suggest that **RBP/RBP4** alters insulin sensitivity in part by affecting insulin signalling in muscle through alterations in the amount of tyrosine-phosphorylated IRS-1 and PI(3)K activation. Thus, RBP/RBP4 may contribute to the pathogenesis of type 2 diabetes, and lowering RBP/RBP4 could be a new strategy for treating type 2 diabetes.

Indications

- Early detection of tubular proteinuria
- Chronic liver diseases
- Cadmium poisoning
- Studies of insulin resistance

3. MATERIAL SUPPLIED

Cat. No.	Label	Kit components	Quantity
K 6120	PLATE	Microtiter plate, pre-coated	12 x 8 wells
K 0001.C.100	WASHBUF	Wash buffer concentrate, 10x	2 x 100 ml
K 6120	SAMPLEBUF	Sample dilution buffer, ready-to-use	1 x 100 ml

Cat. No.	Label	Kit components	Quantity
K 6120	CONJ	Conjugate concentrate (rabbit anti RBP/RBP4, peroxidase-labelled)	1 x 200 µl
K 6120	CAL	Calibrator, lyophilised (see specification for concentration)	2 x 1 vial
K 6120	CTRL 1	Control, lyophilised (see specification for range)	2 x 1 vial
K 6120	CTRL 2	Control, lyophilised (see specification for range)	2 x 1 vial
K 0002.15	SUB	Substrate (tetramethylbenzidine), ready-to-use	1 x 15 ml
K 0003.15	STOP	Stop solution, ready-to-use	1 x 15 ml

For reorders of single components, use the catalogue number followed by the label as product number.

4. MATERIAL REQUIRED BUT NOT SUPPLIED

- Ultrapure water*
- Calibrated precision pipettors and 10–1000 µl single-use tips
- Foil to cover the microtiter plate
- Horizontal microtiter plate shaker
- Multi-channel pipets or repeater pipets
- Centrifuge, 3000 g
- Vortex
- Standard single-use laboratory glass or plastic vials, cups, etc.
- Microtiter plate reader

* Immundiagnostik AG recommends the use of ultrapure water (water type 1; ISO 3696), which is free of undissolved and colloidal ions and organic molecules (free of particles > 0.2 µm) with an electrical conductivity of 0.055 µS/cm at 25 °C (≥ 18.2 MΩ cm).

5. PREPARATION AND STORAGE OF REAGENTS

- To run the assay more than once, ensure that reagents are stored at the conditions stated on the label. **Prepare only the appropriate amount necessary for each run.** The kit can be used up to 4 times within the expiry date stated on the label.
- Reagents with a volume less than **100 µl** should be centrifuged before use to avoid loss of volume.

- **Preparation of the wash buffer:** The **wash buffer concentrate (WASHBUF)** has to be diluted with ultrapure water **1:10** before use (100 ml WASHBUF + 900 ml ultrapure water), mix well. Crystals could occur due to high salt concentration in the concentrate. Before dilution, the crystals have to be redissolved at room temperature or in a water bath at 37°C. The **WASHBUF** is stable at **2–8°C** until the expiry date stated on the label. **Wash buffer** (1:10 diluted WASHBUF) can be stored in a closed flask at **2–8°C for 1 month**.
- Use **100 µl** of **wash buffer** (1:10 diluted WASHBUF) as **blank**.
- The **lyophilised calibrator (CAL)** and **controls (CTRL)** are stable at **2–8°C** until the expiry date stated on the label. Before use, the **CAL** (calibrator) and **CTRL** (controls) must be reconstituted with **500 µl of ultrapure water**. Allow the vial content to dissolve for 10 minutes and mix thoroughly by gentle inversion to ensure complete reconstitution. **Reconstituted calibrators and controls can be stored at 2–8°C for two weeks**.
- The **lyophilised calibrator (CAL)** and **controls (CTRL)** are stable at **2–8°C** until the expiry date stated on the label. Before use, the CAL and CTRL have to be reconstituted with **500 µl of ultrapure water** and mixed by gentle inversion to ensure complete reconstitution. Allow the vial content to dissolve for 10 minutes and then mix thoroughly. **Standards and controls** (reconstituted STD and CTRL) **can be stored at 2–8°C for 2 weeks**.
- **Preparation of the conjugate:** Before use, the **conjugate concentrate (CONJ)** has to be diluted **1:101** in **wash buffer** (100 µl CONJ + 10 ml wash buffer). The **CONJ** is stable at **2–8°C** until the expiry date stated on the label. **Conjugate** (1:101 diluted CONJ) **is not stable and cannot be stored**.
- All other test reagents are ready-to-use. Test reagents are stable until the expiry date (see label) when stored at **2–8°C**.

6. SPECIMEN COLLECTION AND PREPARATION

Plasma and serum

Samples can be stored for two weeks at 4°C. For longer storage, freeze at or below -20°C.

Dilute samples **1:5 000 in sample dilution buffer (SAMPLEBUF)** before use.

- **20 µl** sample + **980 µl** SAMPLEBUF = **1:50 (dilution I)**
- **50 µl** dilution I + **450 µl** SAMPLEBUF = **1:10 (dilution II)**
- **50 µl** dilution II + **450 µl** SAMPLEBUF = **1:10 (dilution III)**

For analysis, pipet **100 µl** of **dilution III** per well.

Urine

Adjust the urine to pH between 6 and 8 with 1 N NaOH. Samples are stable at 2–8 °C for 2 weeks. For longer storage, freeze at or below -20 °C.

Before use, dilute urine **1:10 in sample dilution buffer** (SAMPLEBUF), for example:

100 µl urine + 900 µl SAMPLEBUF, mix well = **1:10**

Urine with an **RBP4 concentration > 330 µg/l must be diluted 1:100**, for example:

10 µl urine + 990 µl SAMPLEBUF, mix well = **1:100**

For analysis, pipet **100 µl** of the **diluted sample** per well.

7. ASSAY PROCEDURE

Principle of the test

This ELISA is designed for the quantitative determination of retinol-binding protein RBP/RBP4 in plasma, serum and urine.

In a first incubation step, RBP/RBP4 in the samples is bound to polyclonal rabbit anti RBP/RBP4 antibodies, immobilised on the microtitre plate. A peroxidase-conjugated anti RBP/RBP4 antibody is used for detection and quantification, and tetramethylbenzidine (TMB) as a peroxidase substrate. The concentration of Retinol-binding protein RBP/RBP4 can be quantified by referring the optical density of the calibrator to a lot-dependent master calibration curve.

Test procedure

Bring all **reagents and samples to room temperature** (15–30 °C) and mix well.

Mark the positions of standards/controls/blank/samples on a protocol sheet.

Take as many microtiter strips as needed from the kit. Store unused strips together with the desiccant bag in the closed aluminium packaging at 2–8 °C. Strips are stable until expiry date stated on the label.

For automated ELISA processors, the given protocol may need to be adjusted according to the specific features of the respective automated platform. For further details please contact your supplier or Immundiagnostik AG.

We recommend to carry out the tests in duplicate.

1.	Before use, wash the wells 5 times with 250 µl wash buffer . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.
2.	Add each 100 µl calibrator/controls/blank/diluted samples into the respective wells.
3.	Incubate for 1 hour at room temperature (15–30°C) on a horizontal shaker* .
4.	Discard the content of each well and wash 5 times with 250 µl wash buffer . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.
5.	Add 100 µl conjugate (diluted CONJ) into each well.
6.	Incubate for 1 hour at room temperature (15–30°C) on a horizontal shaker* .
7.	Discard the content of each well and wash 5 times with 250 µl wash buffer . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.
8.	Add 100 µl substrate (SUB) into each well.
9.	Incubate for 10–20 minutes** at room temperature (15–30°C) in the dark .
10.	Add 100 µl stop solution (STOP) into each well and mix well.
11.	Determine absorption immediately with an ELISA reader at 450 nm against 620 nm (or 690 nm) as a reference. If no reference wavelength is available, read only at 450 nm. If the extinction of the highest standard exceeds the range of the photometer, absorption must be measured immediately at 405 nm against 620 nm as a reference.

* We recommend shaking the strips at 550 rpm with an orbit of 2 mm.

** The intensity of the colour change is temperature sensitive. We recommend observing the colour change and stopping the reaction upon good differentiation.

8. RESULTS

For result evaluation, please use a four parametric logit-log model based on the standard curve of the respective kit lot and the calibrator value (CAL). All essential information on the standard curve is provided on the QC data sheet of the respective product lot.

The calibration curve can be expressed either by the concentration of each standard with its corresponding optical density or by the four parameters A,B,C and D. In both cases the optical density of the calibrator (CAL) is essential. Depending on your evaluation software program, either the one or the other kind of data described above should be entered.

Caution: Please make sure that all parameters and values are transferred accurately into your software as minor deviations can cause severe errors during evaluation.

The plausibility of the duplicate values should be examined before the automatic evaluation of the results. If this option is not available with the programme used, the duplicate values should be evaluated manually.

Serum and plasma

The obtained results have to be multiplied with the **dilution factor of 5 000** to get the actual concentrations.

In case **another dilution factor** has been used, multiply the obtained result with the dilution factor used to get the real concentration.

Urine

The obtained results have to be multiplied with the **dilution factor used** to get the actual concentrations.

9. LIMITATIONS

Samples with concentrations above the measurement range (see definition below) can be further diluted and re-assayed. Please consider this higher dilution when calculating the results.

Samples with concentrations lower than the measurement range (see definition below) cannot be clearly quantified.

The upper limit of the measurement range can be calculated as:

highest concentration of the calibration curve × sample dilution factor to be used

The lower limit of the measurement range can be calculated as:

Analytical sensitivity × sample dilution factor to be used

Analytical sensitivity see chapter "Performance Characteristics".

10. QUALITY CONTROL

Immundiagnostik AG recommends the use of external controls for internal quality control, if possible.

Control samples should be analysed with each run. Results, generated from the analysis of control samples, should be evaluated for acceptability using appropriate statistical methods. The results for the patient samples may not be valid if within the same assay one or more values of the quality control sample are outside the acceptable limits.

Reference range

Plasma or Serum:

Adults	20–75 mg/l
Newborn	11–34 mg/l
Age 6 months	18–50 mg/l

Urine: 0.01–0.54 mg/l

We recommend each laboratory to establish its own reference range.

11. PERFORMANCE CHARACTERISTICS

Dilution recovery

One patient sample was diluted and analysed. The results are shown below (n = 1):

Sample	Dilution	expected [µg/l]	measured [µg/l]
A	1:7000	4.8	4.8
	1:14000	2.8	2.4
	1:28000	1.2	1.2
	1:56000	0.6	0.8

Analytical Sensitivity

The detection limit was set as $B_0 + 2 \text{ SD}$ and estimated to be 0.9 µg/l.

Specificity

No cross reactivity to the carrier protein thyreoglobulin was observed.

Precision and reproducibility

Intra-Assay (n = 16)

The reproducibility of two results in one measurement series was evaluated. Two samples were analysed 16 times by one person using the RBP/RBP4 ELISA.

Sample	RBP/RBP4 mean value [µg/l]	Intra-Assay CV [%]
1	24.1	5
2	11.1	5

Inter-Assay (n = 25)

The reproducibility of two results at different days was evaluated. Two samples were analysed at different days by different persons using the RBP/RBP4 ELISA.

Sample	RBP/RBP4 mean value [µg/l]	Inter-Assay CV [%]
1	4.4	9.8
2	6.9	9.7

12. PRECAUTIONS

- All reagents in the kit package are for *in vitro* diagnostic use only.
- Human materials used in kit components were tested and found to be negative for HIV, Hepatitis B and Hepatitis C. However, for safety reasons, all kit components should be treated as potentially infectious.
- Kit reagents contain sodium azide or Proclin as bactericides. Sodium azide and Proclin are toxic. Substrates for the enzymatic colour reactions are toxic and carcinogenic. Avoid contact with skin or mucous membranes.
- The stop solution consists of diluted sulphuric acid, a strong acid. Although diluted, it still must be handled with care. It can cause burns and should be handled with gloves, eye protection, and appropriate protective clothing. Any spill should be wiped up immediately with copious quantities of water. Do not breath vapour and avoid inhalation.

13. TECHNICAL HINTS

- Do not interchange different lot numbers of any kit component within the same assay. Furthermore we recommend not assembling wells of different microtiter plates for analysis, even if they are of the same batch as wells from already opened microtiter plates are exposed to different conditions than sealed ones.
- Control samples should be analysed with each run.
- Reagents should not be used beyond the expiration date stated on kit label.
- Substrate solution should remain colourless until use.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Avoid foaming when mixing reagents.
- Do not mix plugs and caps from different reagents.
- The assay should always be performed according to the enclosed manual.












14. GENERAL NOTES ON THE TEST AND TEST PROCEDURE

- This assay was produced and distributed according to the IVD guidelines of 98/79/EC.
- Quality control guidelines should be followed.
- Incubation time, incubation temperature and pipetting volumes of the components are defined by the producer. Any variation of the test procedure, which is not coordinated with the producer, may influence the results of the test. Immundiagnostik AG can therefore not be held responsible for any damage resulting from incorrect use.
- Warranty claims and complaints regarding deficiencies must be logged within 14 days after receipt of the product. The product should be send to Immundiagnostik AG along with a written complaint.

15. REFERENCES

1. Graham, T. E., Wason, C. J., Blüher, M. & Kahn, B. B. Shortcomings in methodology complicate measurements of serum retinol binding protein (RBP4) in insulin-resistant human subjects. *Diabetologia* **50**, 814–23 (2007).
2. Graham, T. E. et al. Retinol-binding protein 4 and insulin resistance in lean, obese, and diabetic subjects. *N. Engl. J. Med.* **354**, 2552–63 (2006).
3. Yang, Q. et al. Serum retinol binding protein 4 contributes to insulin resistance in obesity and type 2 diabetes. *Nature* **436**, 356–62 (2005).
4. Blumsohn, A., Morris, B. W., Griffiths, H. & Ramsey, C. F. Stability of β 2-microglobulin and retinol binding protein at different values of pH and temperature in normal and pathological urine. *Clin. Chim. Acta* **195**, 133–137 (1991).
5. Bernard, A. M., Moreau, D. & Lauwerys, R. Comparison of retinol-binding protein and β 2-microglobulin determination in urine for the early detection of tubular proteinuria. *Clin. Chim. Acta* **126**, 1–7 (1982).

Used symbols:

	Temperature limitation		Catalogue Number
	In Vitro Diagnostic Medical Device		To be used with
	Manufacturer		Contains sufficient for <n> tests
	Lot number		Use by
	Attention		Consult instructions for use
	Consult specification data sheet		