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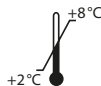
Manual

ID-Vit[®] Biotin

***Microbiological test kit for the determination of biotin in serum
using a Lactobacillus plantarum coated microtitre plate
For use in human and veterinary medicine and in research***

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IVD **CE**



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1. INTENDED USE

ID-Vit® Biotin is a microtiter plate test kit based on a microbiological method which measures the Biotin content in serum. The test kit contains the standard and all reagents required to perform the test. It is sufficient for 96 determinations including standard curves. An ELISA reader is required for the evaluation of the results. For use in human and veterinary medicine and in research. For *in vitro* diagnostic use only.

2. INTRODUCTION

Biotin (vitamin H) is widespread in bacteria, fungi, higher plants and animal tissues. The major part of biotin in food is covalently bound to protein with only a small part being freely available. During digestion, biocytin (biotinyl-lysine) is released from the proteins and can, similarly to biotin, easily be taken up from the intestine. Biotin is then released by the enzyme biocytinase from biocytin in plasma and erythrocytes and is available as prosthetic group for several biotin-dependent enzymes.

The daily biotin requirement is difficult to estimate because a healthy intestinal flora endogenously synthesises biotin and thereby helps to satisfy this need. Recent findings suggest that adults need a daily intake of 100–200 µg biotin. Supplementation on a milligram scale leads to significant improvement regarding neuropathology and glucose metabolism of chronic hemodialysis patients.

Biotin deficiency

Symptoms of biotin deficiency are caused by e.g. damage to the intestinal flora or extreme diets (e.g. frequent consumption of raw eggs). Consequences of biotin deficiency include dermatitis, hair loss, anorexia, muscular hypotonia, depression and sexual dysfunction.

Indications for a determination of biotin

- Defects in enzymes (e.g. genetic deficiency of biotinidase)
- Short gut syndrome
- Altered intestinal flora
- Malnutrition

3. PRINCIPLE OF THE TEST

The serum samples are diluted and then added into the wells of a microtiter plate coated with *Lactobacillus plantarum*. The addition of biotin in either standards or samples gives a biotin-dependent growth response until biotin is consumed. After incubation at **37 °C** for **48 h**, the growth of *Lactobacillus plantarum* is measured turbidimetrically at 610–630 nm (alternatively at 540–550 nm) in an ELISA reader and

compared to a standard curve generated from the dilution series. The amount of biotin is directly proportional to the turbidity.

4. MATERIAL SUPPLIED

Cat. No.	Label	Kit components	Quantity
KIF007	PLATE	<i>Lactobacillus plantarum</i> -precoated microtiter plate	1 x
KIF007	DIL	Water	4x 30 ml
KIF007	ASYMED	Biotin assay medium	4 x
KIF007	STD	Biotin standard, lyoph.	4 x
KIF007	FOL	Adhesive cover foil	4 x
KIF007	FRA	Replacement holder for microtiter strips	1 x
KIF007	CTRL1	Biotin control 1, lyoph.	4 x
KIF007	CTRL2	Biotin control 2, lyoph.	4 x

5. MATERIAL REQUIRED BUT NOT SUPPLIED

- Incubator with a dark incubation chamber, 37 °C
- Water bath (90 °C–100 °C)
- ELISA reader 610–630 nm (540–550 nm)
- Calibrated precision pipettors and sterile 20–1000 µl tips
- 5 ml and 10 ml pipets
- 1.5–2 ml reaction vials, sterile
- 0.2 µm sterile polyethersulfone (PES) filter with a sterile disposable syringe
- 15 ml centrifuge tubes, sterile (e.g. Falcon tubes)
- Biocentrifuge (10 000 g)

6. PRECAUTIONS

- As the test is based on a microbiological method, the general guidelines for sterile work should be observed as far as possible (preferably work in a sterile bench / PCR hood, use of sterile instruments or equipment).
- GLP (Good Laboratory Practice) guidelines have to be observed.

- Water quality is extremely important for the test. Only the water delivered with the test kit [DIL] should be used.
- For sterile filtration, only a sterile polyethersulfone filter must be used.
- It is essential to run a standard curve for each separate assay.
- Controls should be measured with each assay.
- We recommend measurements in duplicate.
- If a higher dilution results in a higher value measured, inhibitors like antibiotics might be present.
- Reagents should not be used beyond the expiration date shown on the label.
- Wear gloves during the test.
- Used microtiter stripes [PLATE] and materials that have been in contact with patient samples should be handled and disposed as potentially infectious.

7. STORAGE AND PREPARATION OF REAGENTS

- Store test kit and reagents at 2–8 °C.
- Prepare reagents freshly and use them immediately after preparation. Discard remaining unused reagents and waste in accordance with country, federal, state, and local regulations.
- To run the assay more than once, ensure that reagents are stored at the conditions stated on the label. Prepare only the appropriate amount necessary for each run. The kit can be used up to 4 times within the expiry date stated on the label.

7.1 Water

- Water [DIL] (for medium [ASYMED], standard [STD] and controls [CTRL1, CTRL2])
- Push the lid up and pull it back to the rim of the glass, then twist the whole cap off.

7.2 Preparation of the controls

- The lyophilised controls [CTRL1, CTRL2] have to be resuspended with each 200 µl water [DIL] from the test kit, then homogenise using a vortex.

- After reconstitution, the controls are treated like samples.
- The concentration of the controls changes from lot to lot and is stated in the product specification.

7.3 Preparation of the standard curve

- For the preparation of the standard curve, standard concentrate is needed. To prepare standard concentrate, resuspend the lyophilised standard [STD] with x ml (x = please see the enclosed quality control protocol for the volume needed) water [DIL] supplied with the test kit, then homogenise using a vortex.
- Prepare a standard curve in 6 sterile reaction tubes (1.5–2 ml volume) from standard concentrate and water [DIL] following the scheme depicted in the table below:

Biotin [µg/l]	Water [DIL] [µl]	+	Standard concentrate [µl]	=	Total volume [µl]
Blank: 0	500	+	0	=	500
Standard 1: 0.02	450	+	50	=	500
Standard 2: 0.06	350	+	150	=	500
Standard 3: 0.10	250	+	250	=	500
Standard 4: 0.14	150	+	350	=	500
Standard 5: 0.18	50	+	450	=	500

7.4 Preparation of the sterile assay medium

- Fresh sterile assay medium has to be prepared each time before performing a test.
- Remove lyophilised assay medium from the desiccant bag in the assay medium bottle by taking the bag with a forceps and shaking it whilst still inside the bottle. Then remove the clean desiccant bag and discard it.
- Add 10 ml water [DIL] to the assay medium bottle [ASYMED], close the bottle firmly and shake it. This amount is sufficient for 6 microtiter stripes.
- Heat the medium bottle in a water bath at 90–100 °C for 5 min, shake well at least 2 times during this incubation time. Take care that the medium bottle is always firmly closed.

- Quickly cool the medium bottle to $< 30^{\circ}\text{C}$ (at $2-8^{\circ}\text{C}$ for 10 min).
- Filter the medium using a disposable syringe (10 ml) and the $0.2\ \mu\text{m}$ PES filter into a sterile centrifuge tube (15 ml, e. g. Falcon).
- After this preparation, the sterile assay medium can be used in the test.

7.5 Microtiter plate [PLATE]

- Store the microtiter plate [PLATE] in the aluminium packaging containing the desiccant bag at $2-8^{\circ}\text{C}$.
- The microtiter plate [PLATE] has to be protected from humidity and contamination.
- Take care that the aluminium packaging is not damaged.
- Carefully close the aluminium packaging after opening.
- Take only the microtiter stripes needed directly before usage to avoid contamination

8. SAMPLE STORAGE AND PREPARATION

- Use serum for analysis.
- Samples are stable at $2-8^{\circ}\text{C}$ for one day in the dark. For longer storage, samples should be frozen and kept at -20°C .
- Hemolytic samples may give erroneous results and should not be used for analysis. Lipemic samples should be centrifuged at $13\ 000g$ before assaying to obtain fat free serum as far as possible.
- Samples should be centrifuged (at least 5 min at $10\ 000g$) prior to measurement. Use the resulting supernatant in the test.

8.1 Sample dilution

Take $50\ \mu\text{l}$ from each sample/control, add $950\ \mu\text{l}$ water [DIL] and mix. The sample dilution results in a total dilution of 1:20 (= sample dilution factor).

9. ASSAY PROCEDURE

9.1 Test preparations

Take as many microtiter strips as needed from kit. Return unused strips and any unused test kit component to the original packaging, and put in the refrigerator. Bring all necessary reagents to room temperature.

9.2 Test procedure

- Take as many microtiter strips as needed from the kit and put them in the second microtiter strip holder [FRA].
- Put 150 µl sterile assay medium into the cavities.
- Add each 150 µl of the prepared standard curve, samples and controls into the respective cavities. Pre-rinse each pipet tip with standard, control or sample solution, respectively.
- Carefully seal the plate with adhesive cover foil [FOL]. Important: the cavities must be made airtight by pressing the foil down with the hand!
- Keep at 37 °C for 48 h in an incubator.

9.3 Measurement

- Press the adhesive cover foil [FOL] firmly down again with the hand.
- Upturn the microtiter plate [PLATE], put it onto a tabletop and shake the microbes well.
- Turn the microtiter plate [PLATE] over again and carefully remove the adhesive cover foil [FOL]. During this, fix the strips in the frame with your hand because the foil is highly adhesive.
- Remove air bubbles in the cavities using a pipet tip or a needle.
- Read turbidity in an ELISA reader at E 610–630 nm (alternatively at E 540–550 nm).

Please note

- After 48 h incubation time, the microtiter plate [PLATE] may be stored for a maximum of 48 h in the refrigerator before measuring the turbidity.
- To prevent time-loss through public holidays or weekends, the microtiter plate [PLATE] may also be evaluated after 60 h incubation.

10. EVALUATION OF RESULTS

We recommend to use the 4 parameter algorithm to calculate the results. The sample dilution factor has to be considered for data evaluation.

The blank should have an optical density < standard 1. It serves as optical control to exclude contaminations and is not included in the calculation of results.

10.1 Calculation

Biotin in $\mu\text{g/l}$ = value from the standard curve \times sample dilution factor (20)

Reference value for human serum

Based on studies of serum samples of apparently healthy persons ($n = 84$), the following biotin values were estimated.

<750 ng/l	biotin deficiency requiring treatment
750–1250 ng/l	suboptimal biotin status
>1250 ng/l	sufficient levels of biotin

Please note

A concentration range of 0.4–3.6 $\mu\text{g/l}$ Biotin is covered at a sample dilution of 1:20.

We recommend each laboratory to develop its own normal range as normal ranges strongly depend on the choice of the patient collective. The values mentioned above are only for orientation and can deviate from other published data.

10.2 Quality control

The extinction of the highest standard has to be > 0.6.

Results, generated from the analysis of control samples, should be evaluated for acceptability. The results for the samples may not be valid if within the same assay one or more values of the quality control sample or the highest standard are outside the acceptable limits.

11. LIMITATIONS

Whole blood cannot be used in the assay.

12. PERFORMANCE CHARACTERISTICS

The following performance characteristics have been collected using human serum samples.

12.1 Precision and reproducibility

Intraassay (n = 5)

	Biotin [$\mu\text{g/l}$]	CV [%]
Sample	63.5	3.4

Interassay (n = 5)

	Biotin [$\mu\text{g/l}$]	CV [%]
Sample	64.3	2.9

12.2 Recovery

Samples from 4 patients were spiked with biotin and analysed. The mean values are shown below.

Sample (n=5)	Mean value original sample [$\mu\text{g/l}$]	Spike [$\mu\text{g/l}$]	Biotin expected [$\mu\text{g/l}$]	Biotin measured [$\mu\text{g/l}$]	Recovery Rate [%]
A	1.1	2	3.1	3.3	110
		4	5.1	5.2	103
		6	7.1	7.3	103
Recovery rate in total [%]					105

Sample (n=5)	Mean value measured in original sample [µg/l]	Spike [µg/l]	Biotin expected [µg/l]	Biotin measured [µg/l]	Recovery Rate [%]
B	1.5	2	3.5	3.2	85
		4	5.5	4.7	80
		6	7.5	7.4	98

Recovery rate in total [%]

88

Sample (n=5)	Mean value measured in original sample [µg/l]	Spike [µg/l]	Biotin expected [µg/l]	Biotin measured [µg/l]	Recovery Rate [%]
C	1.4	2	3.4	3.3	95
		4	5.4	5.6	105
		6	7.4	7.0	93

Recovery rate in total [%]

98

Sample (n=5)	Mean value measured in original sample [µg/l]	Spike [µg/l]	Biotin expected [µg/l]	Biotin measured [µg/l]	Recovery Rate [%]
D	1.3	2	3.3	3.6	115
		4	5.3	5.7	110
		6	7.3	6.9	93

Recovery rate in total [%]

106

12.3 Linearity

Two patient samples were diluted and analysed. The results are shown below:

Sample	Dilution	Biotin expected [µg/l]	Biotin measured [µg/l]
A	20	3.0	3.1
	40		3.5
	80		3.3
	120		2.9
	160		3.4
B	20	1.3	1.3
	40		1.3
	80		1.6

13. REFERENCES












Burtis, C.A. & Ashwood, E.R. eds., Tietz Textbook of Clinical Chemistry 3rd ed., Saunders.

14. GENERAL NOTES ON THE TEST AND TEST PROCEDURE

- This assay was produced and distributed according to the IVD guidelines of 98/79/EC.
- All reagents in the kit package are for *in vitro* diagnostic use only.
- ID-Vit® is a trademark of Immundiagnostik AG.
- Reagents should not be used beyond the expiration date stated on the kit label.
- Do not interchange different lot numbers of any kit component within the same assay.
- The guidelines for medical laboratories should be followed.
- Incubation time, incubation temperature and pipetting volumes of the components are defined by the producer. Any variation of the test procedure, which is not coordinated with the producer, may influence the results of the test. Immundiagnostik AG can therefore not be held responsible for any damage resulting from incorrect use.

- Warranty claims and complaints regarding deficiencies must be logged within 14 days after receipt of the product. The product should be sent to Immunodiagnostik AG along with a written complaint.
- Control samples should be analysed with each run.
- The assay should always be performed according to the enclosed manual.

Used symbols:

	Temperature limitation		Catalogue Number
	In Vitro Diagnostic Medical Device		To be used with
	Manufacturer		Contains sufficient for <n> tests
	Lot number		Use by
	Attention		Consult instructions for use
	Consult specification data sheet		