Maxime PCR PreMix Kit (i-MAX II)

for 20_{µl} rxn

Cat. No. INT-25265 (96 tubes)

DESCRIPTION

Maxime PCR PreMix Kit (i-MAX II) 's characteristics are there. Increased fidelity of PCR amplification due to the i-MAXTM II DNA Polymerase enzyme blend combines the proofreading activity of Pfu DNA Polymerase with the high processivity of Taq DNA Polymerase. The second is that increased yield of PCR amplification. Finally, improved performance of long PCR because the reaction buffer and the enzyme blend are optimized for generation of certain length products. So, it can amplify even longer fragments up to 17.5kb from human genomic DNA, and up to 30kb from a DNA template. Maxime PCR PreMix (i-MAX II) is the product what is mixed every component: i-MAX™ II DNA Polymerase, dNTP mixture, reaction buffer, and so on- in one tube for 1 rxn PCR. This is the product that can get the best result with the most convenience system. The first reason is that it has every components for PCR, so we can do PCR just add a template DNA, primer set, and D.W.. The second reason is that it has Gel loading buffer to do electrophoresis, so we can do gel loading without any treatment. In addition, each batches are checked by a thorough Q.C., so its reappearance is high. It is suitable for various sample's experience by fast and simple using method.

STORAGE

Store at -20°C; under this condition, it is stable for at least a year.

CHARACTERISTICS

- · Increased fidelity of PCR amplification
- · Increased yield of PCR amplification
- : Because the accuracy of the enzyme blend reduces the number of truncated amplification products formed.
- · Improved performance of long PCR
- · Ready to use: only template and primers are needed
- Stable for over 1 year at -20 °C
- · Time-saving and cost-effective

CONTENTS

Maxime PCR PreMix (i-MAX II; for 20 μl rxn)

96 tubes.

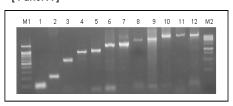
Component in 20 µl reaction					
⊬MAX™ (II) DNA Polymerase	2.5 U				
dNTPs	2.5 mM each				
Reaction Buffer	1x				
Gel Loading buffer	1x				

Note: The PCR process is covered by patents issued and applicable in certain countries. iNtRON Biotechnology does not encourage or support the unauthorized or unlicensed use of the PCR process. Use of this product is recommended for persons that either have a license to perform PCR or are not required to obtain a license.

EXPERIMENTAL INFORMATION

· Amplification of various templates 87 bp to 20 Kb.

[Panel A]



[Panel B]						
М1	1	2	3	4	M2	
=					0	
			-			
Ξ	_					
_						
			16			

Extension

Final extension

Figure 1. Amplification of various template 97bp to 20Kb with Maxime PCR PreMix Kit (i-Max II)

[Panel A] Using various template

Lane M1, 100bp Ladder DNA Marker; lane M2, 1Kb Ladder DNA Marker; lane 1, 87bp; lane 2,200bp; lane 3, 570bp, lane 4, 1Kb; lane 5, 1.3Kb; lane 6, 1.8kb; lane 7, 2Kb; lane 8, 2.7Kb; lane 9, 4.5Kb; lane 10, 9Kb; lane 11, 17.5Kb; lane 12, 20Kb

[Panel B] Using only human gDNA template

Lane M1, 1Kb Ladder DNA Marker; lane M2, \(\lambda \text{Hind III Marker; lane 1, 1.8Kp; } \) lane 2, 2Kb; lane 3, 2.7Kb, lane 4, 17.5Kb

PROTOCOL

- 1. Add template DNA and primers into *Maxime* PCR PreMix tubes (¿MAX II). Note 1: Recommended volume of template and primer: 3µl~5µl
- 2. Add distilled water into the tubes to a total volume of 20µl. Do not calculate the dried components.

	Example	Example Total 20μl reaction volume			
•	PCR reaction r	nixture	Add		
•	Template DNA	(I pg ~ 1 μg)	1 ~ 2μΙ		
	Primer (F : 5 ~	10pmol/μl)	1μΙ		
	Primer (R : 5 ~	10pmol/μl)	1μΙ		
_	Distilled Water		16 ~ 17μΙ		
	Total reaction	volume	20 μl		

Note: This example serves as a guideline for PCR amplification. Optimal reaction conditions such as amount of template DNA and amount of primer, may vary and must be individually determined.

3. Dissolve the blue pellet by pipetting

Cycle program for fragments < 10kb

Note: If the mixture lets stand at RT for 1-2min after adding water, the pellet is easily dissolved.

4. (Option) Add mineral oil.

Note: This step is unnecessary when using a thermal cycler that employs a top heating method(general methods).

- 5. Perform PCR of samples.
- 6. Load samples on agarose gel without adding a loading-dye buffer and perform electrophoresis.

CYCLING PARAMETERS FOR SHORT & LONG FRAGMENTS

	Temp.	Time	Cycle No.			
Initial Denaturation	92-94℃	2-4min	1			
Denaturation Annealing Extension*	94 ℃ 45-65 ℃ 72 ℃	15s-1min 15s-1min 1min/1-1.5kb	25-30			
Final extension	72℃ 4℃	5-10min hold	1			
*, Extension time for 30s-1min is sufficient for fragments < 1kb.						
Cycle program for fragments > 10kb						
	Temp.	Time	Cycle No.			
Initial Denaturation	92-94℃	2-4min	1			
Denaturation Annealing Extension	94 ℃ 45-65 ℃ 72 ℃	15s-1min 15s-1min 1min/1-1.5kb	10			
Denaturation Annealing Extension	94 ℃ 45-65 ℃	15s-1min 15s-1min 1min/1-1 5kh				

Note: This CYCLING PARAMETERS serves as a guideline for PCR amplification. optimal reaction conditions such as PCR cycles, annealing temperature, extension temperature and incubation times, may vary and must be individually determined.

72°C

72°C



1min/1-1.5kb

+ 20s/cycle

5-10min